



# Use of RAPD to Develop Markers for Identifying Buartnut Hybrids between butternut and Japanese walnut

Peng Zhao<sup>1</sup> Keith Woeste<sup>2</sup>

<sup>1</sup> College of Forestry, Northwest Agriculture and Forestry University, Yangling, Shaanxi, 712100, China

<sup>2</sup> Hardwood Tree Improvement and Regeneration Center, Department of Forestry and Natural Resources, Purdue University, West Lafayette, IN 47907, USA



## Abstract

The survival of butternut (*Juglans cinerea* L.), a temperate hardwood, is threatened by butternut canker, a disease incited by the exotic fungus *Sirococcus clavignenti juglandacearum*. Field observations indicate that the hybrid known as buartnut [a cross of butternut and its close congener Japanese walnut (*J. cinerea* × *J. ailantifolia*)] may be more resistant to butternut canker than is either parental species. Scientists have expressed concern over the possibility of range-wide genetic invasion by Japanese walnut (Ostry & Woeste 2004). Unfortunately, hybrids are often difficult to distinguish from butternuts. Pair wise combinations forty random primers were used to screen a panel of genotypes of butternut, Japanese walnut and buartnuts to identify genomic region unique to Japanese Walnut. About 530 randomly amplified polymorphic DNA (RAPD) panels were examined. We have found about twenty DNA amplicons present in Japanese walnut and buartnut hybrids but absent in butternut. We have cloned nine of these markers in preparation for sequencing. These markers will be used to identify buartnut hybrids based on the presence of introgressed genomic fragments inherited from Japanese walnut.

## Introduction

Butternut (*Juglans cinerea* L.) also called white walnut, lemonsut, or oilnut is a native, short-lived, cold-tolerant tree species formerly valued for its nuts, wood and wildlife mast. Butternut is native to eastern North America, from New Brunswick, south to Georgia and west to Minnesota and Arkansas (Rink 1990). The nuts are usually used in baking and making candies. Butternut is threatened by butternut canker, a disease incited by the exotic fungus *Sirococcus clavignenti juglandacearum*. Butternut canker infects and kills butternut trees throughout the natural range of the species (Orchard 1984). Japanese walnut (*Juglans ailantifolia* Carr.), native to Japan and Sakhalin was introduced into America from Japan about 1870 by a nurseryman at San Jose, California. Hybrids between butternut and Japanese walnut are known as Buartnut (technically *Juglans* × *bixbyi*) are often more resistant to butternut canker than either parental species. Unlike most *Juglans* hybrids, buartnuts are highly fruitful and vigorous, and they are able to cross with other hybrids, both parental species, and may even self-pollinate, producing trees with confusing combinations of traits. Biologists have expressed concern over the possibility of range-wide genetic invasion by Japanese walnut (Ostry & Woeste 2004). Unfortunately, hybrids are often difficult to distinguish from butternuts. Differences in protein mobility or DNA sequence among members of the Juglandaceae have become a mainstay of *Juglans* phylogenetics and conservation genetics (Germain et al. 1993, Fjellstrom and Parfitt 1995, Stanford et al. 2000, Orel et al. 2003, Aradhya et al. 2007, Ross-Davis and Woeste 2007). Although most of these studies included both butternut and Japanese walnut genotypes, there are only a few species-specific markers for these taxa (Allozyme, Germain et al. 1993, Ross Davis et al. 2008). These markers are already being used to identify non-hybrid trees in National Forests for use in establishing seed orchards and to further butternut breeding efforts by the Forest Service and public cooperating institutions. Our objective was to use of RAPD (Randomly Amplified Polymorphic DNAs) to develop markers for identifying butternut, Japanese walnut and buartnut hybrids.

## Materials and Methods

Leaf samples were harvested from germ plasma maintained by the Hardwood Tree Improvement and Regeneration Center (HTIRC) at Purdue University. Vouchered specimen *Juglans ailantifolia* were obtained from the National Clonal Germplasm Repository Davis, CA. DNA was extracted from leaf samples using methods of Robichaud (1997) and stored at -80 °C. DNA concentration was estimated using a Nanotrop (ND-1000) and the DNA concentration was diluted with TLE buffer. DNA quality was evaluated electrophoretically. Bulk DNA samples screened with RAPD (Randomly Amplified Polymorphic DNAs) primers from Gene Link (Hawthorne, NY). We screened over 500 primers combinations for amplicons present in Japanese walnut and buartnut but absent from butternut (Table 1). Forty promising primers or primer combinations were chosen for this study (Table 2). PCR reactions contained 1X Taq reaction buffer (50mM), MgCl<sub>2</sub> (0.4mM), dNTP(0.25mM), Bovine Serum Albumin, acetylated (0.1mg ml<sup>-1</sup>), and primers (5μM). The final reaction was 20μl, including Zui sample DNA (1.0 to 0ng, μl<sup>-1</sup>) and 0.5 units Taq polymerase. Thermal cycling conditions were as follows: denaturation 3 min at 92°C; 35 cycles of 1 min at 92 °C, 1 min at 35 °C and 2 min at 72 °C; and final extension 10 min at 72 °C. DNA bands were extracted from agarose gel with QIAquick Gel Extraction Kit. DNA was ligated onto pCEM-T and Pgm-T Easy Vectors. We used electroporation to transform high efficiency competent cells. Transformed colonies were identified using blue and white selection add ampicillin. Plasmid DNA was extracted with Zippy™ Plasmid Miniprep Kit.

Table 1 40 RAPD decamer primers Used for screening Buartnut hybrids

Primer Name	Sequence (5' to 3')	Size (bp)	Present in JA	Present in JB	Present in JC
20A	CGAGGCTCT	282	+	+	-
20B	TTGCGGCTG	282	+	+	-
40A	AAATGAGAT	282	+	+	-
40B	AAATGAGAT	282	+	+	-
40C	AAATGAGAT	282	+	+	-
40D	AAATGAGAT	282	+	+	-
40E	AAATGAGAT	282	+	+	-
40F	AAATGAGAT	282	+	+	-
40G	AAATGAGAT	282	+	+	-
40H	AAATGAGAT	282	+	+	-
40I	AAATGAGAT	282	+	+	-
40J	AAATGAGAT	282	+	+	-
40K	AAATGAGAT	282	+	+	-
40L	AAATGAGAT	282	+	+	-
40M	AAATGAGAT	282	+	+	-
40N	AAATGAGAT	282	+	+	-
40O	AAATGAGAT	282	+	+	-
40P	AAATGAGAT	282	+	+	-
40Q	AAATGAGAT	282	+	+	-
40R	AAATGAGAT	282	+	+	-
40S	AAATGAGAT	282	+	+	-
40T	AAATGAGAT	282	+	+	-
40U	AAATGAGAT	282	+	+	-
40V	AAATGAGAT	282	+	+	-
40W	AAATGAGAT	282	+	+	-
40X	AAATGAGAT	282	+	+	-
40Y	AAATGAGAT	282	+	+	-
40Z	AAATGAGAT	282	+	+	-

## Results

Figure 1 RAPD Panels Showing No Difference Between Butternut (JC), Japanese Walnut (JA) and Buartnut (JB)

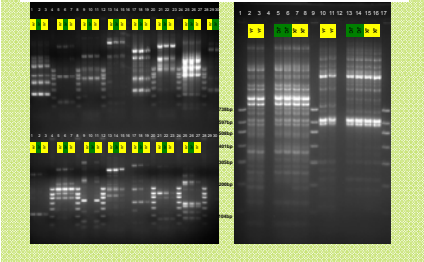


Figure 2 Primer Combination A1-B4

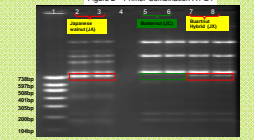


Figure 3 Primer Combination A1-B4

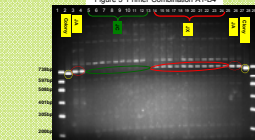


Figure 4 Primer Combination B20-A13

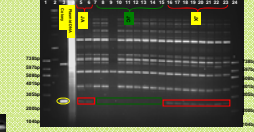


Figure 5 Primer Combination B20-A13

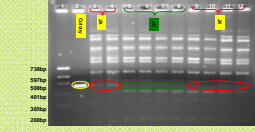


Figure 6 Primer Combination B15-B10

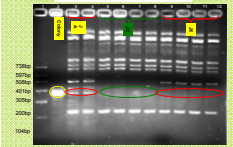


Figure 7 Primer Combination B15-B2



Figure 8 Primer Combination B20-A13

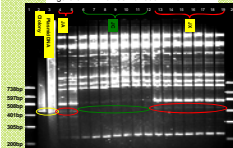


Figure 9 Two Examples of Cloned RAPD Amplicons

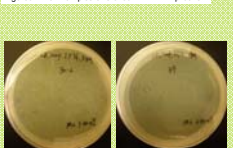


Table 2 Candidate Amplicons Present in JA and JB but Absent from JC

Primer	DNA Size	Checked against population	Cloned	Primer	DNA Size	Checked against population	Cloned
A1B4	738bp	✓	✓	B19A10	150bp	✓	✓
B18A16	220bp	✓	✓	B20A7	210bp	✓	✓
B20A13	490bp	✓	✓	B20A7	750bp	✓	✓
B20A13	410bp	✓	✓	B20A8	520bp	✓	✓
B15B4	260bp	✓	✓	B20A8	500bp	✓	✓
B20A8	420bp	✓	✓	B19B5	680bp	✓	✓
B20A8	360bp	✓	✓	B19B8	750bp	✓	✓
B15B6	450bp	✓	✓	B19B10	800bp	✓	✓
B15B8	490bp	✓	✓	B13A13	470bp	✓	✓
B15B10	470bp	✓	✓	B11A8	590bp	✓	✓
B15B12	260bp	✓	✓	B11A9	210bp	✓	✓
B11B12	260bp	✓	✓	B12A8	420bp	✓	✓
B20A13	320bp	✓	✓	B12A8	380bp	✓	✓
B20A13	410bp	✓	✓	B12A9	420bp	✓	✓
B20A8	280bp	✓	✓	B12A11	380bp	✓	✓
B14B12	260bp	✓	✓	A10B1	538bp	✓	✓
B12A8	420bp	✓	✓	B15B10	230bp	✓	✓
B15B10	220bp	✓	✓	B15B12	150bp	✓	✓
B12A9	320bp	✓	✓	B12A2	250bp	✓	✓
B13A13	470bp	✓	✓	A19B11	220bp	✓	✓
B15B2	320bp	✓	✓	A19B11	350bp	✓	✓

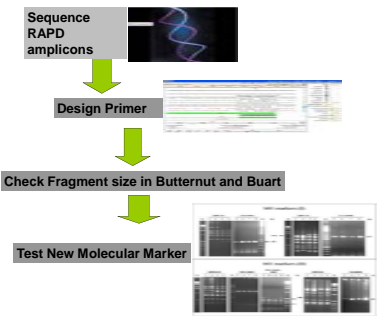
## Literature cited

Aradhya, M.K., D. Potter, F. Gao and C.J. Simon. 2007. Molecular phylogeny of *Juglans* (Juglandaceae): a biogeographic perspective. *Tree Genet. Genom.* 3:363-378.  
Bronzi de Caratta Virginia, Jean Gnanetier, Claude Garbottti & Jacques Masny. Genetic relationships between cultivated and wild olives of Corsica and Sardinia using RAPD markers *Euphytica* 123: 263-271, 2002  
Faxon, K. Woeste, L. W. Monti, R. Rao. Identification of Sonoran walnut using simple sequence repeats (SSRs). *Genet. Resour. Crop Evol.* (2007) 54: 1981-1984. DOI: 10.1007/s10722-006-9187-0  
Germain, E., I. Hanauer & R. Moret. 1993. Identification of eight *Juglans* spp. and their interspecific hybrids by isozymic electrophoresis. *Acta Horticulturae* 311: 73-81.  
Gomez V. J., A. D. Marchant, M. C. Tamayo, R. Medina, A. Fernandez P. (2004) Genetic diversity in the olive tree (*Olea europaea* L.) using microsatellite markers. *Genet. Resour. Crop Evol.* 51:301-311  
Gould S. D., Keith Woeste, et al. Characterization of 14 Microsatellite Markers for Genetic Analysis of Walnut. *J. Amer. Soc. Hort. Sci.* 133(3): 348-354, 2008.  
Fjellstrom, R.G. and C.E. Parfitt. 1995. Phylogenetic analysis and evolution of the genus *Juglans* (Juglandaceae) as determined from nuclear genome RFLPs. *Plant Syst. Evol.* 107:19-32.  
Hoban Sean M., Tim G. McClean, Scott E. Schlarbaum and Jacques Romo-Servano. Geographically extensive hybridization between the forest trees American butternut and Japanese walnut. *Biol. Lett.* doi:10.1098/rstb.2009.0031  
Nose, F.P., J.I. Hormaza & G.H. McCreanran. Molecular characterization and genetic relationships among walnut (*Juglans regia* L.) genotypes based on RAPD markers. *Euphytica* 101: 199-206, 1998  
Orel, G., A.D. Marchant, J.A. McLeod and G.D. Robinson. 2003. Characterization of 11 Juglandaceae genotypes based on microsatellite markers. *Genet. Resour. Crop Evol.* 50:176-183.  
Osby, M. E., Woeste, K. Sonnet of butternut canker in North America, host range, evidence of resistance within butternut populations and conservation genetics. *Proceeding of the 6th Walnut Council Research Symposium*, 2004.  
Reh, C. 1990. *Juglans cinerea* L., butternut. In *Silvics of North America*, Vol. 2. Hardwoods, Eds. R.M. Burns and B.W. Honninger. USDA For. Serv., Washington DC. Agric. Handbook 381:176-183.  
Robichaud R. L., C. Claubitz, O.E. Rhodes, Jr., and K. Woeste. A robust set of black walnut microsatellites for parentage and clonal identification. *New Forests* 2006, 32:179-196  
Ross-Davis, A. and K.E. Woeste. 2007. Microsatellite markers for *Juglans cinerea* L. and their utility in other Juglandaceae. *Cons. Genet.* <http://dx.doi.org/10.1007/s10542-006-9046-4>  
Stanford, A., R. Hansen and C.P. Parks. 2000. Phylogeny and biogeography of *Juglans* (Juglandaceae) based on mark and 17S rDNA sequence data. *Am. J. Bot.* 87:872-882.  
Woeste, K., G.H. McCreanran and R. Benarthy. 1995. Randomly amplified polymorphic DNA loci from a walnut backcross (*Juglans hindsii* × *J. regia*). *J. Am. Soc. Hort. Sci.* 1996 121:359-361.

## Conclusion

Pair wise combinations of forty random primers were used to screen a panel of genotypes of butternut, Japanese walnut and buartnuts to identify genomic region unique to Japanese Walnut. About 530 randomly amplified polymorphic DNA (RAPD) panels were examined. We have found about twenty DNA amplicons present in Japanese walnut and buartnut hybrids but absent in butternut. We have cloned nine of these markers in preparation for sequencing to identify markers that distinguish butternut and hybrid genotypes. The markers were derived from RAPD primers A1B4, B12B13, B15B8, B15B6, B15B10, B15B2, B20A8 and B20A13. By sequencing these amplicons we expect to develop markers with less complex amplification products that can be used to identify hybrids (buartnut) and butternut trees for use in establishing seed orchards and to further butternut breeding efforts by the Forest Service and public cooperating institutions. These markers will be used to identify buartnut hybrids based on the presence of introgressed genomic fragments inherited from Japanese walnut.

## Ongoing and Future Work



## Acknowledgements

I would like to thank my advisor Dr. Woeste. I am grateful to the Department of Forestry and Natural Resources and the Hardwood Tree Improvement and Regeneration Center for the opportunity to perform my Ph.D. research. Thanks to my lab mates in the laboratories of Dr. Woeste and Marcia Kremer for purchasing everything for my research, and my fellow students: Zhonglian Huang, Lisa Worthen, Chad Shultz, Hannah Bergeman and Ningxia Du for their support and help. I thank my family, including my wife, parents and siblings, for their understanding during my study and research.